

Inside Diagnostics

FECAL CALPROTECTIN - A NEW TEST FOR THE ASSESSMENT OF INTESTINAL INFLAMMATION

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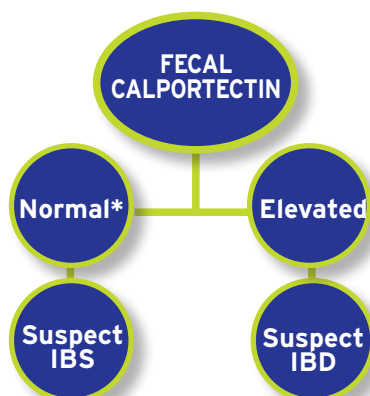
Inflammatory bowel disease (IBD) and irritable bowel syndrome (IBS) are conditions that present with chronic abdominal pain and disrupted bowel habits. Combined, they affect more than 5 million Canadians indiscriminately of age, gender or race¹.

To Scope or Not to Scope

Aside from negatively impacting the quality of life, IBS is relatively benign. It does not lead to any permanent structural damage to the intestines. Dietary and lifestyle changes can usually alleviate the symptoms of this disorder². IBD on the other hand is characterized by intestinal inflammation and ulcers. This organic disease can lead to serious complications including the development of fistulas, abscesses and strictures; sometimes leading to perforation of the bowel³. This disease also carries an increased risk of malignancy⁴. Diagnosing IBD typically involves endoscopy and medical imaging which is expensive, invasive and difficult to conduct, especially with young patients.

Helping You Diagnose and Monitor IBD

Calprotectin may be used to confirm the presence of intestinal inflammation before using invasive testing⁵. Fecal Calprotectin is an FDA approved test that offers a non-invasive way to assess for the presence of intestinal inflammation. Calprotectin is a calcium-binding protein found within neutrophils in cases with bowel inflammation. Fecal Calprotectin is detected via an immunoassay of a stool sample. The levels of the protein are high in cases of IBD but not IBS, therefore this test helps differentiate between the two disorders.



* Repeat testing for borderline result is recommended.

CLINICAL UTILITY OF FECAL CALPROTECTIN

- Differentiate between IBS and IBD
- Monitor the effectiveness of IBD therapy
- Detect IBD relapse

While Fecal Calprotectin is not a substitute for other diagnostic techniques, it could help identify the patients who are most likely affected by IBD⁵.

The Fecal Calprotectin test is also appropriate for monitoring of the patients who are being treated for IBD⁶. Elevated levels of Calprotectin could be indicative of disease flare-up, thereby allowing physicians to intervene before the IBD symptoms become severe.

IMPORTANT NOTES

- **To avoid potential false positive results, patients should abstain from using NSAIDs for at least two weeks before taking the test⁷.**
- **Repeat testing for borderline results is recommended.**

If you would like to receive more information about this test or order Fecal Calprotectin pamphlets for your patients, please email LifeLabs at contactus@lifelabs.com.

REFERENCES

1. *Digestive Disorders*. Oakville, ON: Canadian Digestive Health Foundation, n.d. Web. 11 Jan 2013.
2. *Irritable Bowel Syndrome*. Bethesda, MD: The National Digestive Diseases Information Clearinghouse. NIH Publication (2007) 07-693.
3. Carty E, et al. Evaluation of new therapies for inflammatory bowel disease. *Br J Clin Pharmacol* (2003) 56(4): 351.
4. Bernstein CN, et al. Cancer risk in patients with inflammatory bowel disease: a population-based study. *Cancer* (2001) 15; 91(4):854.
5. Sherwood RA. Faecal Markers of Gastrointestinal Inflammation. *J Clin Pathol* (2012) 65(11): 981.
6. Vermeire G, et al. Laboratory Markers In IBD: Useful, Magic, Or Unnecessary Toys? *Gut* (2006) 55: 426.
7. Carroccio A, et al. Diagnostic Accuracy of Fecal Calprotectin Assay in Distinguishing Organic Causes of Chronic Diarrhea from Irritable Bowel Syndrome: A Prospective Study in Adults and Children. *Clin Chem* (2003) 49(6): 861.

ROLE OF AMYLASE AND LIPASE TESTING IN PANCREATITIS

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Pancreatitis is an inflammation caused by the injury of pancreatic acinar cells due to activation of digestive enzymes within the pancreatic tissue and is characterized by significant morbidity and mortality.^{1,2} Traditionally, pancreatitis has been classified as:

- a) **Acute pancreatitis:** A reversible inflammation due to enzymatic necrosis, which leads to acute onset of persistent epigastric and/or periumbilical pain, nausea and vomiting.^{1,3} The most common causes of acute pancreatitis include obstruction of the common bile duct by stones (38%) and alcohol abuse (36% of patients). In rare cases acute pancreatitis can be caused by abdominal surgery, extreme hyperlipidemia (triglyceride concentrations > 11 mmol/l),

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congenital anomalies (pancreas divisum), drugs (e.g. azathioprine, steroids, valproate, furosemide), and viral infections (e.g. mumps).

b) Chronic pancreatitis: A progressive inflammation with irregular fibrosis, duct dilation, and irreversible loss of pancreatic parenchyma.^{2,4} It presents either as recurrent attacks mimicking acute pancreatitis, or as a constant, disabling abdominal pain. It is often associated with maldigestion and steatorrhea due to pancreatic insufficiency with loss of enzymes, and glucose intolerance or diabetes due to islet damage. Most commonly the cause of chronic pancreatitis is unknown (60-70%), whereas the most common known cause is alcohol (30% of patients). Other, rare causes include drugs (valproate, thiazide), hyperlipidemia, infections (HIV, mumps), autoimmune, genetic (cystic fibrosis), obstruction (cancer), and recurrent acute pancreatitis.

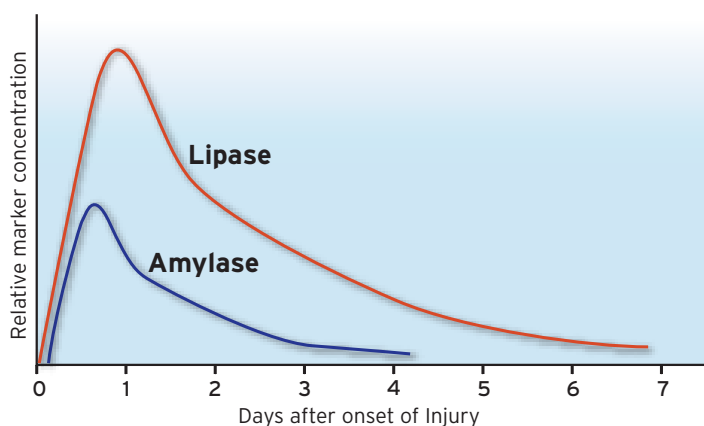
More recently, acute pancreatitis, recurrent attacks and chronic pancreatitis have been regarded as a disease continuum.⁴ Due to different underlying pathophysiological processes, the utility of the laboratory and imaging tests varies throughout the disease continuum. This article summarizes the advantages and limitations of testing amylase, lipase or their combination in pancreatitis.

What Diagnostic Tests are Available to Aid Diagnosis of Acute Pancreatitis?

The non-specific clinical picture of acute pancreatitis represents a diagnostic challenge. Laboratory and imaging studies are necessary for prompt and accurate diagnosis, to expedite appropriate treatment and reduce the risk of complications, including the early organ dysfunction.^{1,3}

Laboratory testing can confirm the diagnosis of acute pancreatitis. The laboratory tests most commonly used are serum amylase and lipase – pancreatic enzymes released into blood following autodigestion of pancreatic tissue during the inflammatory process.^{1,3,5} Blood levels of both enzymes increase markedly within 2-8 hours of symptom onset (Figure 1) and elevations greater than three times the upper limit of normal are typically considered diagnostic.¹ Assays for both enzymes are readily available in the laboratory.

Figure 1. Time-dependent changes in serum amylase and lipase after acute pancreatitis (adapted from reference 5).



Imaging studies to ascertain clinical diagnosis have found that serum amylase has a 81 – 95% sensitivity^{3,6}. Factors decreasing amylase sensitivity include late patient presentation as serum amylase levels normalize 2-4 days after symptom onset, (Figure 1), and normal amylase levels reported in up to 30% of patients with alcoholic pancreatitis and other exocrine pancreatic insufficiencies, and in up to 50% of patients with simultaneous hypertriglyceridemia (the latter due to an unidentified circulatory inhibitor of amylase in hypertriglyceridemic serum).⁶ The major limitation of serum amylase is its lack of specificity.^{5,6} High concentrations of amylase are secreted from salivary glands, and it may also increase in a number of abdominal and extra-abdominal conditions (Table 1).

Table 1. Conditions Associated with Increased Serum Amylase and Lipase^{3,7}

Amylase	Lipase
1) Intra-abdominal: <ul style="list-style-type: none"> • Pancreatic: acute pancreatitis, pseudocysts, trauma, cancer • Non-pancreatic: appendicitis, hepatitis, cholecystitis, peptic ulcer, perforated bowel, mesenteric ischemia, intestinal obstruction, peritonitis, abdominal aortic aneurysm, ruptured ectopic pregnancy, fallopian and ovarian cysts, salpingitis. 	1) Intra-abdominal <ul style="list-style-type: none"> • Pancreatic: acute pancreatitis, pseudocysts, trauma, cancer • Non-pancreatic: cholecystitis, peptic ulcer, mesenteric ischemia, abdominal trauma
2) Extra-abdominal: salivary diseases, decreased glomerular filtration, ketoacidosis, pneumonia, cerebral trauma, burns, anorexia nervosa, bulimia, non-abdominal surgery	2) Extra-abdominal: bone fracture, fat embolism
3) Macroamylasemia*	3) Macrolipasemia*
4) Idiopathic hyperamylasemia: familial, non-familial	
5) Drug-induced: azathioprine, sulfonamides, tetracycline, methyl dopa, estrogens, furosemide, valproic acid, salicylate, thiazide, calcium	

* Asymptomatic presence of macromolecular complexes of enzyme and immunoglobulins

Recent studies have shown that lipase demonstrates better clinical performance compared to amylase, with sensitivity of 85-100% that does not decrease with late patient presentation, as serum lipase remains elevated for up to 14 days following symptom onset (Figure 1).^{3,6} Lipase also performs better in patients with alcoholic pancreatitis and hypertriglyceridemia.⁶ Lipase has better specificity than amylase, but small amounts are present in stomach, intestine, leukocytes, fat cells and milk. Thus, lipase may also be elevated in several conditions other than acute pancreatitis (Table 1). Finally, multiple isoforms of lipase present in serum may interfere with testing.³

Studies directly comparing amylase and lipase for the diagnosis of acute pancreatitis have demonstrated that lipase has superior diagnostic accuracy on both day one and day three following symptom onset.⁸⁻¹⁰ Furthermore, when both enzymes are tested simultaneously, there is no significant improvement in diagnostic accuracy.^{9,11} There is no advantage in serial enzyme measurement, as neither amylase nor lipase levels are related to disease severity or prognosis.^{1,3}

What About Amylase and Lipase in Chronic Pancreatitis?

The loss of pancreatic parenchyma in chronic pancreatitis leads to the loss of enzymes and pancreatic insufficiency.⁴ Although elevations might exist during the acute pain episodes in the early course of the disease, pancreatic amylase and lipase are typically normal or low in advanced chronic pancreatitis.² Thus, monitoring these enzymes in patients with chronic disease does not provide useful clinical information. The treatment goals in chronic pancreatitis, including calming the disease process and relieving pain, correction of metabolic abnormalities, such as diabetes and malnutrition, and management of complications, will dictate a different battery of laboratory tests needed to monitor the patient.

Conclusion

Table 2 summarizes advantages and limitations of enzyme testing in acute pancreatitis. When suspecting acute pancreatitis, it is prudent to take into account the timing of symptom onset, as well as other aspects of the clinical picture. Serum amylase and/or lipase are not useful for monitoring patients with chronic pancreatitis.

Table 2. Advantages and Limitations of Testing Amylase, Lipase or Both for Diagnosis of Acute Pancreatitis

Test	Advantages	Limitations
Amylase	<ul style="list-style-type: none"> • Sensitive in early diagnosis • Readily available 	<ul style="list-style-type: none"> • Lower sensitivity in late presenting patients, alcoholic pancreatitis and hypertriglyceridemia • Low specificity • No standardized method • Different diagnostic cut-offs used
Lipase	<ul style="list-style-type: none"> • Sensitive in both early and late diagnosis • Better specificity compared to amylase • Readily available 	<ul style="list-style-type: none"> • No standardized method, technically more difficult method • Different diagnostic cut-offs used
Amylase + Lipase	None	<ul style="list-style-type: none"> • No improvement in diagnostic accuracy compared to lipase • Increased cost compared to single enzyme testing

POINTS TO REMEMBER

- Lipase is more sensitive and specific than amylase in diagnosing acute pancreatitis.
- Lipase shows better diagnostic accuracy for acute pancreatitis compared to amylase, particularly in late presenting patients, patients with alcoholic pancreatitis and other forms of pancreatic insufficiency, and patients with hypertriglyceridemia.
- Ordering both tests provides no significant improvement in diagnostic accuracy for acute pancreatitis.
- There is no role for serial monitoring of amylase and/or lipase in patients with chronic pancreatitis.

REFERENCES

1. Wu BU, Conwell DL, Banks PA. Chapter 25. Acute Pancreatitis. In: Greenberger NJ, Blumberg RS, Burakoff R, eds. *CURRENT Diagnosis & Treatment: Gastroenterology, Hepatology, & Endoscopy*. 2nd ed. New York: McGraw-Hill; 2012. <http://www.accessmedicine.com/content.aspx?aID=55958678>. Accessed February 14, 2013.
2. Conwell DL, Wu B, Banks PA. Chapter 26. Chronic Pancreatitis. In: Greenberger NJ, Blumberg RS, Burakoff R, eds. *CURRENT Diagnosis & Treatment: Gastroenterology, Hepatology, & Endoscopy*. 2nd ed. New York: McGraw-Hill; 2012. <http://www.accessmedicine.com/content.aspx?aID=55958824>. Accessed February 14, 2013.
3. Frossard J-L, Steer ML, Pastor CM. Acute pancreatitis. *Lancet* 2008;371:143.
4. Braganza JM, Lee SH, McCloy RF, McMahon MJ. Chronic Pancreatitis. *Lancet* 2011;377:1184.
5. Panteghini M, Bais R. Chapter 22. Serum Enzymes. In: Burtis CA, Ashwood ER, Bruns DE, eds. *Tietz Textbook of Clinical Chemistry and Molecular Diagnostics*. 5th ed. St Louis, MO: Elsevier Saunders, 2012.
6. Yadav D, Agarwal N, Pitchumoni CS. A critical evaluation of laboratory tests in acute pancreatitis. *Am J Gastroenterol* 2002;97:1309.
7. Harper SJE, Cheslyn-Curtis S. Acute pancreatitis. *Ann Clin Biochem* 2011;48:23-27.
8. Treacy J, Williams A, Bais R, et al. Evaluation of amylase and lipase in the diagnosis of acute pancreatitis. *ANZ J Surg* 2001;71:577.
9. Keim V, Teich N, Fiedler F, et al. A comparison of lipase and amylase in the diagnosis of acute pancreatitis in patients with abdominal pain. *Pancreas* 1998;16:45.
10. Smith RC, Southwell-Keely J, Chesher D. Should serum pancreatic lipase replace serum amylase as a biomarker of acute pancreatitis? *ANZ J Surg* 2005;75:3999.
11. Werner M, Steinberg WM, Pauley C. Strategic use of individual and combined enzyme indicators for acute pancreatitis analyzed by receiver-operator characteristics. *Clin Chem* 1989;35:967.

OHIP REQUISITION - ESSENTIAL INFORMATION

Quality and Regulatory Affairs LifeLabs ON

MOHLTC regulation mandates community laboratories obtain written requests from qualified healthcare practitioners before any tests are performed. Healthcare practitioners have an obligation to provide a signed and legible OHIP requisition to their patients. The information on the OHIP requisition will ensure tests are ordered and reported in a timely manner. Healthcare practitioners may access the essential information that must be provided for the laboratory to perform the analysis on LifeLabs website www.lifelabs.com. Look for: 'Healthcare Clients-Ontario' Click on: *Laboratory Services* and then *OHIP Requisition Essential Information* folder (see illustration).

For healthcare practitioners who collect specimens in their facility, LifeLabs will provide a package of information and charts that will assist in collection and submission of quality specimens.

As an Ontario Laboratory Accreditation (OLA) accredited laboratory LifeLabs also requires the phlebotomist who has collected the specimens in the facility, to record their initials on the requisition in the top left hand corner.

LifeLabs OHIP Requisition Essential Information

MOHLTC Requisition Essential Information NOTE: Separate requisitions are required for cytology, histology, pathology and tests performed by Public Health Laboratory

LEGEND

1. Phlebotomist initials
2. Ordering Client's name and address
3. Ordering Client's billing number
4. Reporting requirements (i.e. fax)
5. Any pertinent clinical information
6. Copy of Client's FULL NAME AND ADDRESS (phone number) where ordering Client can be reached, including after hours number
7. Date of Service (yyyy-mm-dd)
8. Patient's current health card number
9. Patient's current version code
10. Patient's sex
11. Patient's date of birth (yyyy-mm-dd)
12. Patient's phone number
13. Patient's last name
14. Patient's first and middle names
15. Patient's address

1. Tests ordered (indicating testing vs. random when required)
2. MUST BE CLEAR AND LEGIBLE
3. Profile terminology cannot be used - individual tests must be listed separately
4. Indicate source
5. Indicate whether PISA or VIL. C is insured or uninsured
6. Time and date of last dose for therapeutic drugs
7. Time of collection (when applicable - therapeutic drug/monitoring collection) (24 hr clock)
8. Date of collection (yyyy-mm-dd)
9. Signature of ordering Client or authorized delegate and date signed

Ensure initials of Phlebotomist are recorded on the requisition

OLA Feb 2013

POINTS TO REMEMBER

- Separate requisitions are needed for cytology, pathology and tests performed by the Public Health Laboratory.

GASTROINTESTINAL PARASITIC INFECTIONS

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Although more common in the developing world, parasitic infections of the intestinal tract remain an important clinical entity in Canada due to a number of factors, including increased international travel to developing countries, increasing numbers of immunocompromised individuals because of chemotherapy, solid organ or hematological transplants, increased crowding in day care centers and nursing homes, consumption of raw or partially cooked foods and contaminated water sources, and increased immigration from developing countries. Parasitic pathogens as etiologic agents of diarrhea in developed countries occur with different frequencies in immunocompetent and immunocompromised patients.

Common Etiologic Agents of Parasitic Intestinal Infections

- **Cryptosporidium** — Cryptosporidium is one of the most common parasitic causes of acute foodborne diarrhea in North America accounting for 8 percent of cases in a 2005 survey from FoodNet.¹ Infection with *Cryptosporidium* species may present as a severe dehydrating but self-limited diarrheal illness in immunocompetent hosts. In immunocompromised hosts it may have a more prolonged and severe course.

Transmission of cryptosporidiosis occurs via spread from an infected person or animal, or from a fecally contaminated environment such as a food or water source. Numerous waterborne community outbreaks have occurred from contaminated drinking or swimming water.² Person-to-person transmission is also common, particularly among household members, sexual partners, children in daycare centers and their caretakers, and healthcare workers.

- **Giardia** — *Giardia lamblia* is a flagellated protozoan parasite that is one of the two most common gastrointestinal parasites in North America (Cryptosporidium is the other). *G. lamblia* causes both epidemic and sporadic disease and is an important cause of waterborne and foodborne diarrhea in day-care centers.
- **Cyclospora** — *Cyclospora cayetanensis* has been identified in both sporadic cases and as a cause of outbreaks of diarrhea with the source of infection identified, for example, imported fruit or vegetables. The clinical course of diarrhea is often lengthy, with symptoms lasting a number of weeks, and can be accompanied by fatigue and malaise.
- **Entamoeba histolytica** — Intestinal amebiasis is caused by *Entamoeba histolytica*. Amebiasis is a worldwide disease, but developing countries have significantly higher prevalence rates because of poorer socioeconomic conditions and sanitation levels. In developed countries amebiasis is mainly seen in immigrants from and travelers to endemic countries. Institutionalized patients and men who have sex with men are also at increased risk of infection.

Clinical infection usually has a subacute onset, usually over one to three weeks. Symptoms range from mild diarrhea to severe dysentery producing abdominal pain and bloody diarrhea.

When to Send Stool for Testing for Ova and Parasites

There are several possible indications for sending stool for testing for ova and parasites:³

1. Persistent diarrhea (associated with Giardia, Cryptosporidium, and *Entamoeba histolytica*)
2. Persistent diarrhea following travel
3. Persistent diarrhea with exposure to infants in daycare centers (associated with Giardia and Cryptosporidium)
4. Diarrhea in a man who has sex with men (MSM associated with Giardia and *Entamoeba histolytica*) or a patient with HIV infection or other risk factors causing immunosuppression (associated with a variety of parasitic pathogens)
5. A community waterborne outbreak (associated with Giardia and Cryptosporidium)
6. Bloody diarrhea (associated with intestinal amebiasis).

Specimen Collection, Processing and Reporting

If after one stool specimen no parasites are seen, up to three specimens should be sent on consecutive days for parasite examination since parasite excretion may be intermittent in contrast to bacterial pathogens. Stool samples should be taken prior to starting any anti-parasitic treatment. Ingestion of mineral oil, bismuth, or barium should be avoided prior to stool collection as they may interfere with recovery of parasitic pathogens.⁴ Freshly passed stool is collected into a container with SAF preservative, and should be transported to the lab at room temperature within 72 hours of collection.

From the preserved specimen, microscopic examination is done on a concentrated sample and on a direct smear. Fecal concentration is performed for detection of protozoan cysts, helminth eggs and larvae in small numbers. A Kinyoun Haematoxylin stain is performed for detection of protozoan cysts and trophozoites.⁵

Once specimens are examined, any helminth eggs or larvae are reported as are any pathogenic protozoa that are detected. The pathogenic protozoa of clinical importance include those listed above (*Entamoeba histolytica*, *Giardia lamblia*, *Cryptosporidium species*, *Cyclospora cayetanensis*), along with *Microsporidia*, *Dientamoeba fragilis*, *Balantidium coli* and *Isospora belli*.⁶

Of note, it is not possible to microscopically distinguish between pathogenic *E. histolytica* and the non-pathogenic *E. dispar*, therefore when seen, the organism is reported as *Entamoeba histolytic/dispar*. If further testing is needed, an unpreserved stool specimen should be submitted in a sterile container and antigen testing to distinguish between *E. histolytica* and *E. dispar* can be done at the Public Health laboratory.

Overall, parasitic pathogens remain an important etiology of clinically important intestinal symptoms and proper testing and examination of stool specimens is paramount in ensuring recovery and detection of these pathogens.

POINTS TO REMEMBER

- Up to three stool samples taken on consecutive days should be examined for the presence of ova and parasites.
- Stool must be collected and transported to the lab in a container with SAF preservative.
- Some of the organisms with potential to cause clinically important diarrhea which may require treatment include *Entamoeba histolytica*, *Giardia lamblia*, *Cryptosporidium species*, *Cyclospora cayetanensis*, *Microsporidia*, *Dientamoeba fragilis*, *Balantidium coli* and *Isospora belli*.
- On initial examination, the pathogenic *E. histolytica* and the non-pathogenic *E. dispar* cannot be distinguished, therefore if further testing is required, antigen testing can be done at the Public Health Laboratory on an unpreserved stool specimen.

REFERENCES

1. Centers for Disease Control and Prevention (CDC). Preliminary FoodNet data on the incidence of infection with pathogens transmitted commonly through food. *MMWR Morbidity & Mortality Weekly Report* 2006; 55(14):392.
2. Centers for Disease Control and Prevention (CDC). Syndromic surveillance. Reports from a national conference. *MMWR Morbidity & Mortality Weekly Rep.* 2004;53 Suppl:1.
3. Thielman NM, Guerrant RL. Clinical practice. Acute infectious diarrhea. *New England Journal of Medicine* 2004; 350:38.
4. Garcia L.S. *Diagnostic Medical Parasitology*, 5th edition 2007. ASM Press, Washington DC.
5. Isenberg H. *Clinical Microbiology Procedures Handbook*, 3rd edition, ASM Press, 2010.
6. LifeLabs Standard Operating Procedure, Document # 3804. *Intestinal Ova and Parasites*. Revised May 2012

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