COLLECTION OF FINE NEEDLE ASPIRATION (FNA) SPECIMENS

Specimen Labeling:

All specimens will be clearly labeled BEFORE being sent to the laboratory for testing, to ensure correct identification of the patient and sample.

All specimens/containers must be labeled with:

- The patient’s full name (printed in the same format as patient’s health card)
- A second identifier such as date of birth or health card number

It is recommended that the specimen container also be labeled with specimen source (e.g., Left Breast)

Specimen/container labeling options are:

- Computer printed label affixed to the side of the specimen container, or
- Clearly printed handwritten information on the label of the specimen container using indelible ink

Specimen Handling and Transportation:

- Specimens collected from multiple sites must be collected in separate containers with the source of each identified.
- Each specimen must be placed into a polybag.
- A completed Cytology & HPV Testing Requisition must accompany each specimen.
- Specimens requiring expedited service must be clearly marked as such. The typical designation is: ASAP.
- For optimal results transport the specimens to the laboratory as soon as possible after collection.
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Cytology Requisition Information:
All specimens must be submitted for testing with a completed Cytology & HPV Testing Requisition.
Provide the following information in a legible format:

Patient Information:
- Full name of patient (printed in the same format as patient’s health card)
- Health card number
- Date of birth
- Date of specimen collection
- Specimen source
- Anatomic site
- Number specimens submitted (e.g., slides, vials)
- Collection method (e.g., fine needle aspiration)
- Pertinent clinical information

Health Care Provider Information:
- Full name, address and billing number of the ordering health care provider
- Full name, address (and billing number if known) of any copy-to physicians

Collection Kit Information:
Fine Needle/ Sputum/ Urine Kits:
Kit components are ordered separately:
- 90 mL sterile container with 40 mL of cytology preservative (CytoLYT® clear, colourless solution)
- Polybag
- Cytology requisition
(Do not use formalin for the collection of Cytology Specimens)

CAUTION: The preservative contains methyl alcohol. Do not drink. If ingested, do not induce vomiting; call a doctor or local poison control center immediately. Vapor may be harmful if inhaled; use with adequate ventilation. Flammable; keep away from heat, sparks & open flame. Avoid contact with eyes.
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Physician Collection Instructions

The following instructions apply to most palpable superficial lesions. Supplementary instructions for thyroid and salivary FNAB follow.

1. Using standard FNAB technique, sample the area of interest.
2. Expel the contents of the needle barrel into the cytology preservative. For most lesions all of the aspirated material should be placed into Cytolyt fixative. See supplementary method below for thyroid and salivary FNABs.
3. Rinse the needle: Aspirate approximately 2cc of preservative into the syringe, through the needle, to rinse the needle and syringe of any remaining specimen. Express into the specimen container.
4. Usually the first pass yields the most diagnostic material. Diagnostic yield can be improved by using up to 3 passes. The use of more three passes does not generally increase the yield.
5. Tightly re-cap the specimen container.
6. Ensure the specimen container(s), any slide(s), and the requisition are labeled with the patients’ full name and DOB or Health card number, the date of collection, the sample source and site as well as clinical history/impression (see above instructions).
7. Keep the preserved specimen at room temperature or refrigerated (2-8°C). It is recommended to return the specimen to the laboratory as soon as possible after collection. Specimen integrity is compromised after 8 days after collection.
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Supplementary Method: for thyroid and salivary FNAB:

(Aspirates from sites other than thyroid and salivary do not require smears)

- In order to better assess architecture, it is recommended that only a single pair of smeared slides be produced from the first pass and then the needle rinsed into Cytolyt solution. Additional passes should be expelled into the needle rinse.
- Label the frosted end of the slides with the patient/sample information. **Note:** Specimen collected on a glass slide must have the patient/sample information printed on the frosted end of the slide using pencil or indelible ink.
- To prepare the slides – after the first pass, place one drop of aspirated material in the centre of the slide close the the labelled end. With the long edge of a second slide held at 90° smear the material along the slide length.

- Fix both slides immediately with Cytospray, holding the bottle no closer than 6 inches from the slide.
- Expel the remaining aspirated material into the Cytolyt fixative container and rinse the needle. It is important that the majority of the material be fixed in Cytolyt rather than on the smear.
- Further passes should be placed directly into Cytolyt as further smeared slides are unnecessary.
- Allow the smeared slides to dry completely before securely placing into the cardboard/plastic folder for transport.

Reference:
NCCLS GP23-A  Nongynecologic Cytologic Specimens: Collection and Cytopreparatory Techniques; Approved Guideline. Volume 17, Number 19, 1999